

STUDIES ON CEPHALOSPORIN ANTIBIOTICS

I. SYNTHESIS, ANTIBACTERIAL ACTIVITY AND ORAL ABSORPTION OF NEW 3-(*O*-SUBSTITUTED)-7 β -[*D*- α -AMINO- α -(4-HYDROXYPHENYL)ACETAMIDO]CEPHALOSPORINS

CHIHIRO YOKOO, MASAMI GOI, AKIRA ONODERA, MITSUO MURATA,
TAKATOSHI NAGATE, YOSHIKI WATANABE and KAORU SOTA

Research Center, Taisho Pharmaceutical Co., Ltd.,
1-403 Yoshino-cho, Ohmiya, Saitama 330, Japan

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The synthesis, antibacterial activity and oral absorption of new 7 β -[*D*- α -amino- α -(4-hydroxyphenyl)acetamido]cephalosporins (**1**) with various *O*-substituents at the C-3 position of a cephalosporin nucleus are described. Of these, the cephalosporins (**1b**~**1e**) having an alkoxycarbonylmethoxy group at the C-3 position showed good oral absorption in rats as well as potent activity against Gram-positive bacteria. The structure-activity relationships of **1** are also presented.

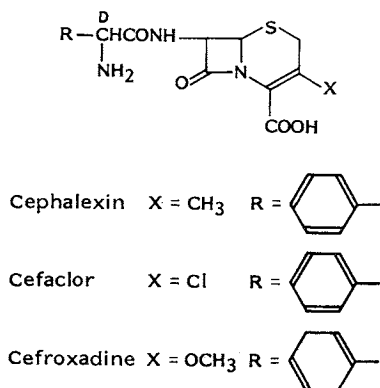
Since cephalixin¹⁾ has been introduced into clinical medicine as an orally active cephalosporin, much research²⁻⁷⁾ has been reported aimed at obtaining new cephalixin analogues with improved properties. In recent years, the new analogues such as cefaclor⁸⁾ and cefroxadine⁷⁾ bearing an electron-negative hetero-atom directly attached to the C-3 position of the cephem nucleus have been developed (Fig. 1).

They are more active than cephalixin, and structurally unique among the many cephalosporins in that no carbon atom is attached to the C-3 position. In order to find more active cephalixin analogues, we also planned to prepare the new derivatives, represented by general structure **1** (Fig. 2), with various *O*-substituents directly attached to the C-3 position.

In this paper, we wish to report the synthesis of **1**, and the effects of the new substituents at the C-3 position on antibacterial activity and oral absorption in rats.

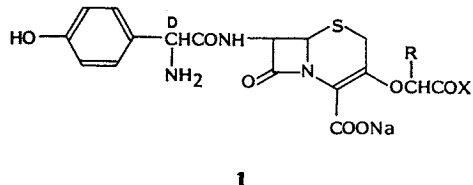
Chemistry

Fig. 1.

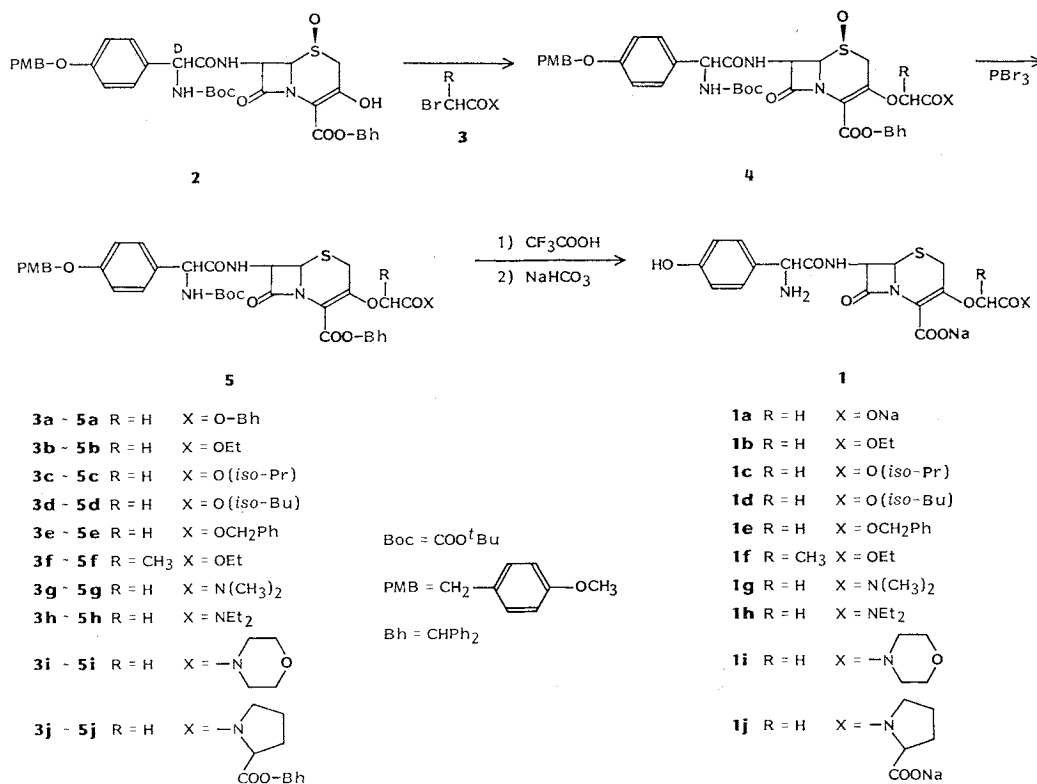


The new cephalosporins (**1a**~**1j**) were synthesized by the route as outlined in Scheme 1. The hydroxy group of diphenylmethyl 7-aryl-glycylamido-3-hydroxycephalosporinate 1-oxide (**2**) prepared by the method shown in Scheme 2

Fig. 2.



Scheme 1.



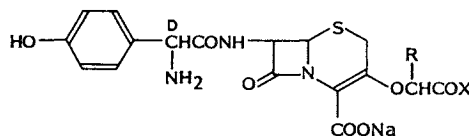
was reacted with 2-bromoacetic and 2-bromopropionic acid derivatives (**3a-3j**) in the presence of *N,N*-diisopropylethylamine as a base to yield the *C*-3 *O*-substituted derivatives (**4a-4j**). The alternative route to **4** with the corresponding diazo compounds such as ethyl diazoacetate in the presence of rhodium (II) acetate⁹ did not proceed smoothly. Subsequently, the sulfoxides (**4a-4j**) were reduced using phosphorus tribromide (PBr_3) in DMF to yield the cephem compounds (**5a-5j**). Removal of the protecting *tert*-butoxycarbonyl (Boc), *p*-methoxybenzyl (PMB) and diphenylmethyl (Bh) groups of **5a-5j** by treatment with trifluoroacetic acid and anisole afforded the new cephem compounds (**1a-1j**).

The common intermediate (**2**) for **1a-1j** was prepared according to the reaction sequence shown in Scheme 2. Diphenylmethyl 7 β -amino-3-(1-methyltetrazol-5-yl)thiomethyl cephalosporinate (**7**)⁹ was acylated with *N*-Boc-4-(4-methoxybenzyl)oxy-D-phenylglycine (**6**) by using *N,N'*-dicyclohexylcarbodiimide (DCC) as a condensing agent. The *C*-7 acylamino compound (**8**) was then reacted with *m*-chloroperbenzoic acid (mCPBA) to give the corresponding sulfoxide (**9**), which was treated with Zn and formic acid¹⁰ to yield the *C*-3 exomethylenecephem compound (**10**).



The ozonolysis of **10** afforded the desired intermediate (**2**). The *C*-7 acyl moiety (**6**) was also prepared starting from *N*-Boc-4-hydroxy-D-phenylglycine (**11**) by conventional methods shown in Scheme 3. In order to prevent undesirable side-reactions in the following reactions, the hydroxy group of **11** was protected with a *p*-methoxybenzyl group¹¹.

Antibacterial Activity and Oral Absorption

The *in vitro* antibacterial activities of the new cephalixin analogues (**1**) against selected Gram-

Table 1. *In vitro* antibacterial activity and peak serum level of 1.

1

No.	Compound		MIC ($\mu\text{g/ml}$, 10^8 cfu/ml) ^a					Peak serum level ($\mu\text{g/ml}$) ^b po, 50 mg/kg, rats ($n=3$)
	X	R	<i>S.a.</i>	<i>S.e.</i>	<i>E.c.</i>	<i>K.p.</i>	<i>P.m.</i>	
1a	ONa	H	100	>100	50	50	50	<3.2
1b	OEt	H	0.78	1.56	6.25	6.25	12.5	42.4
1c	O(<i>iso</i> -Pr)	H	1.56	3.13	50	25	50	31.6
1d	O(<i>iso</i> -Bu)	H	0.78	1.56	25	12.5	100	37.8
1e	OCH ₂ Ph	H	1.56	1.56	50	25	100	30.9
1f	OEt	CH ₃	1.56	1.56	100	50	100	3.6
1g	N(CH ₃) ₂	H	0.78	1.56	12.5	6.25	25	<4.0
1h	NEt ₂	H	1.56	1.56	25	12.5	50	<3.5
1i		H	0.78	1.56	12.5	12.5	50	1.6
1j		H	100	>100	25	25	50	<2.8
Cephalexin			0.78	0.78	12.5	6.25	25	13.3

^a The MICs were determined by a standard agar dilution method using sensitivity test agar (Eiken, Japan).

^b The peak serum levels were measured by a disc-plate method using *Escherichia coli* SC 507 or *Micrococcus luteus* NIHJ as the test organism.

Abbreviations: *S.a.*; *Staphylococcus aureus* 209P JC-1, *S.e.*; *Staphylococcus epidermidis* sp-al-1, *E.c.*; *Escherichia coli* NIHJ JC-2, *K.p.*; *Klebsiella pneumoniae* IFO 3317, *P.m.*; *Proteus mirabilis* IFO 3849.

at the C-3 position plays an important role in the oral absorption.

In this study, we found some new cephalosporin analogues with improved oral absorption in rats by the chemical modification of the C-3 position.

Experimental

MP was determined with a Yanagimoto micro melting point apparatus and are uncorrected. IR spectra were taken on a Jasco DS-701G IR spectrometer. ¹H NMR spectra were recorded on a Varian XL-200 NMR spectrometer using TMS or sodium trimethylsilyl propionate-*d*₄ (in D₂O) as an internal standard. MS was measured on a Jeol JMS-DX303 mass spectrometer. The following abbreviations are used: s, singlet; d, doublet; t, triplet; m, multiplet; br s, broad singlet; ABq, AB quartet.

Determination of Antibacterial Activity

MIC was determined by the agar dilution method using sensitivity test agar (Eiken, Japan) after incubation at 37°C for 18 hours with inoculum size of 10⁸ cfu/ml.

Oral Absorption Study

Male SLC/Wistar rats ($n=3$) weighing 180~220 g were fasted overnight and orally dosed with 50 mg/kg of the test compounds. Serum samples were collected at 0.5, 1, 2 and 4 hours respectively after dosing. The serum levels of the test compounds were measured by the disc-plate method using

Escherichia coli SC 507 or *Micrococcus luteus* NIHJ as the test organism and the sensitivity test agar (Eiken, Japan) as the test medium.

4-Methoxybenzyl *N*-(*tert*-Butoxycarbonyl)-4-(4-methoxybenzyl)oxy-D-phenylglycinate (12)

To a solution of *N*-(*tert*-butoxycarbonyl)-4-hydroxyphenylglycine (11) (25.0 g, 93.5 mm) in acetone (70 ml) were added 4-methoxybenzyl chloride (36.8 g, 235 mm), potassium iodide (31.2 g, 188 mm) and potassium carbonate (26 g, 188 mm) at room temp. After being stirred for 16 hours at the same temp, the reaction mixture was concentrated under reduced pressure to dryness. To the residue, H₂O (200 ml) was added, and extracted with EtOAc (300 ml). The extract was washed with brine (200 ml), dried (MgSO₄) and the solvent was evaporated.

The residue was purified by column chromatography on silica gel (eluent; benzene - acetone, 40 : 1), and crystallized from MeOH to give 41.6 g (87.7%) of 12: MP 72~74°C. IR (KBr) cm⁻¹ 1740, 1705, 1610; ¹H NMR (CDCl₃) δ 1.43 (9H, s, *tert*-Bu), 3.79 (3H, s, OCH₃), 3.81 (3H, s, OCH₃), 4.97 (2H, s, CH₂O), 5.05 and 5.13 (2H, ABq, *J*=11 Hz, COOCH₂), 5.28 (1H, d, *J*=8 Hz, CH(NH)COO), 5.50 (1H, d, *J*=8 Hz, NHCOOC₄H₉), 6.83 (2H, d, *J*=9 Hz, aromatic H), 6.91 (2H, d, *J*=9 Hz, aromatic H), 6.93 (2H, *J*=9 Hz, aromatic H), 7.17 (2H, d, *J*=9 Hz, aromatic H), 7.25 (2H, d, *J*=9 Hz, aromatic H), 7.35 (2H, d, *J*=9 Hz, aromatic H); field desorption mass spectrum (FD-MS) *m/z* 508 (M+1)⁺;

Anal Calcd for C₂₈H₃₃NO₇: C 68.62, H 6.55, N 2.76.

Found: C 68.66, H 6.58, N 2.66.

N-(*tert*-Butoxycarbonyl)-4-(4-methoxybenzyl)oxy-D-phenylglycine (6)

To a solution of 12 (39.6 g, 78.1 mm) in acetone (213 ml) was added 1 N NaOH solution (93.7 ml, 93.7 mm) under ice-cooling, and stirred for 30 minutes at room temp. After removal of acetone under reduced pressure, the resulting aqueous solution was adjusted to pH 2.0 with 0.5% HCl, and extracted with EtOAc (300 ml). The extract was washed with brine (200 ml), dried (MgSO₄) and evaporated to give a crystalline residue, which was collected and washed with Et₂O (100 ml) to afford 25.4 g (84.0%) of 6. Recrystallization from MeOH gave a pure material: MP 145~146°C (dec); IR (KBr) cm⁻¹ 1745, 1675, 1610; ¹H NMR (DMSO-*d*₆) δ 1.37 (9H, s, *tert*-Bu), 3.75 (3H, s, OCH₃), 5.01 (2H, s, CH₂O), 5.03 (1H, d, *J*=8 Hz, CH(NH)COOH), 6.95 (2H, d, *J*=8 Hz, aromatic H), 6.96 (2H, d, *J*=8 Hz, aromatic H), 7.31 (2H, d, *J*=8 Hz, aromatic H), 7.38 (2H, d, *J*=8 Hz, aromatic H), 7.48 (1H, d, *J*=8 Hz, NHCOOC₄H₉), 12.61 (1H, br s, COOH); FD-MS *m/z* 387 (M⁺);

Anal Calcd for C₂₁H₂₅NO₆: C 65.10, H 6.50, N 3.62.

Found: C 65.16, H 6.58, N 3.44.

Diphenylmethyl 7β-[D-α-(*tert*-Butoxycarbonylamino)-α-[4-(4-methoxybenzyl)oxyphenyl]acetamido]-3-(1-methyltetrazol-5-yl)thiomethyl-3-cephem-4-carboxylate (8)

To a solution of diphenylmethyl 7β-amino-3-(1-methyltetrazol-5-yl)thiomethyl-3-cephem-4-carboxylate (7)⁹⁾ (10.0 g, 20.2 mm) in THF (20 ml), were added 6 (8.58 g, 22.2 mm) and DCC (4.57 g, 22.2 mm) under ice-cooling. After being stirred for 3.5 hours at the same temp, the precipitate of *N,N'*-dicyclohexylurea formed was filtered off and the filtrate was concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (eluent; benzene - acetone, 10 : 1), and crystallized from MeOH to give 18.0 g of 8: MP 154~156°C; IR (KBr) cm⁻¹ 1780, 1705, 1660; ¹H NMR (CDCl₃) δ 1.42 (9H, s, *tert*-Bu), 3.69 (2H, br s, 2-H₂), 3.82 (3H, s, OCH₃), 3.84 (3H, s, NCH₃), 4.25 and 4.39 (2H, ABq, *J*=14 Hz, 3'-H₂), 4.97 (1H, d, *J*=5 Hz, 6-H), 4.99 (2H, s, CH₂O), 5.14 (1H, d, *J*=6 Hz, CH(NH)CO), 5.52 (1H, d, *J*=6 Hz, NHCOOC₄H₉), 5.87 (1H, dd, *J*=5 and 9 Hz, 7-H), 6.48 (1H, d, *J*=9 Hz, CONH), 6.92 (2H, d, *J*=8 Hz, aromatic H), 6.95 (1H, s, CHPh₂), 6.97 (2H, d, *J*=8 Hz, aromatic H), 7.23~7.49 (14H, m, aromatic H); FD-MS *m/z* 863 (M⁺);

Anal Calcd for C₄₄H₄₅N₇O₈S₂: C 61.16, H 5.25, N 11.35.

Found: C 61.25, H 5.19, N 11.42.

Diphenylmethyl 7β-[D-α-(*tert*-Butoxycarbonylamino)-α-[4-(4-methoxybenzyl)oxyphenyl]acetamido]-3-(1-methyltetrazol-5-yl)thiomethyl-3-cephem-4-carboxylate 1β-Oxide (9)

To a solution of 8 (18.0 g, 20.8 mm) in CH₂Cl₂ (200 ml) was added mCPBA (3.16 g, 20.9 mm)

under ice-cooling. After being stirred for 30 minutes at the same temp, the reaction mixture was washed with 5% NaHCO₃ (100 ml), brine (100 ml), and dried (MgSO₄). The solvent was evaporated, and the residue was purified by column chromatography on silica gel (eluent; benzene - acetone, 10:1~8:1), and then crystallized from MeOH to give 14.3 g (80.5% from 7) of **9**. Recrystallization from MeOH - CHCl₃ gave a pure material: MP 139~141°C; IR (KBr) cm⁻¹ 1795, 1715, 1690, 1610; ¹H NMR (CDCl₃) δ 1.42 (9H, s, *tert*-Bu), 3.50 (1H, d, *J*=18 Hz, 2-H_α), 3.82 (6H, s, NCH₃ and OCH₃), 3.69 (1H, d, *J*=18 Hz, 2-H_β), 4.09 and 4.55 (2H, ABq, *J*=13 Hz, 3'-H₂), 4.43 (1H, d, *J*=5 Hz, 6-H), 4.98 (2H, s, CH₂O), 5.13 (1H, d, *J*=5 Hz, CH(NH)CO), 5.58 (1H, d, *J*=5 Hz, NHCOOC₄H₉), 6.06 (1H, dd, *J*=5 and 9 Hz, 7-H), 6.92 (2H, d, *J*=8 Hz, aromatic H), 6.94 (1H, s, CHPh₂), 6.96 (2H, d, *J*=8 Hz, aromatic H), 7.24~7.54 (15H, m, aromatic H and CONH); FD-MS *m/z* 880 (M+1)⁺;

Anal Calcd for C₄₄H₄₅N₇O₉S₂: C 60.05, H 5.15, N 11.14.

Found: C 59.76, H 5.10, N 11.13.

Diphenylmethyl 7β-[D-α-(*tert*-Butoxycarbonylamino)-α-[4-(4-methoxybenzyl)oxyphenyl]acetamido]-3-methylenecepham-4-carboxylate 1β-Oxide (**10**)

To a mixture of **9** (14.3 g, 16.3 mm) in THF (94.5 ml) and DMF (27.3 ml) were added Zn dust (11.6 g, 178 mm), HCOOH (27.3 ml) and H₂O (27.3 ml) under ice-cooling. After being stirred for 1 hour at the same temp, the spent Zn was filtrated and washed with EtOAc (200 ml). The separated organic layer was washed with 5% NaHCO₃ (100 ml×2), brine (100 ml) and dried (MgSO₄). The solvent was evaporated, and the residue was purified by column chromatography on silica gel (eluent; CHCl₃ - MeOH, 200:1~100:1) to give 9.3 g (74.5%) of **10** as a white powder: MP 104~108°C; IR (KBr) cm⁻¹ 1780, 1735, 1700, 1685; ¹H NMR (CDCl₃) δ 1.42 (9H, s, *tert*-Bu), 3.32 (1H, d, *J*=14 Hz, 2-H_α), 3.66 (1H, d, *J*=14 Hz, 2-H_β), 3.80 (3H, s, OCH₃), 4.72 (1H, d, *J*=5 Hz, 6-H), 4.96 (2H, s, CH₂O), 5.14 (1H, d, *J*=5 Hz, CH(NH)CO), 5.34 (1H, s, vinyl H), 5.43 (1H, br s, 4-H), 5.56 (1H, d, *J*=5 Hz, NHCOOC₄H₉), 5.81 (1H, s, vinyl H), 5.86 (1H, dd, *J*=5 and 10 Hz, 7-H), 6.86 (2H, d, *J*=8 Hz, aromatic H), 6.92 (1H, s, CHPh₂), 6.94 (2H, d, *J*=8 Hz, aromatic H), 7.22~7.38 (14H, m, aromatic H), 7.63 (1H, d, *J*=10 Hz, CONH); FD-MS *m/z* 765 (M⁺);

Anal Calcd for C₄₂H₄₃N₈O₉S: C 65.86, H 5.66, N 5.49.

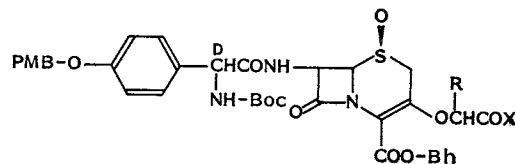
Found: C 65.81, H 5.79, N 5.65.

Diphenylmethyl 7β-[D-α-(*tert*-Butoxycarbonylamino)-α-[4-(4-methoxybenzyl)oxyphenyl]acetamido]-3-hydroxy-3-cephem-4-carboxylate 1β-Oxide (**2**)

Excess ozone was passed through a mixture of **10** (13.9 g, 18.2 mm) in CH₂Cl₂ (1,200 ml) and MeOH (2.5 ml) for 1.5 hours at -40~-30°C until the solution became blue. After removing excess ozone by passing dry nitrogen, dimethyl sulfide (11.2 ml) was added to the mixture at -40°C. The temp of the mixture was slowly raised to 20°C over 1 hour. The resulting solution was washed with brine (300 ml), dried (MgSO₄) and evaporated. The residue was purified by column chromatography on silica gel (eluent; CHCl₃ - MeOH, 50:1) to give 9.82 g (70.3%) of **2** as an amorphous solid. IR (KBr) cm⁻¹ 1785, 1685, 1610; ¹H NMR (CDCl₃) δ 1.46 (9H, s, *tert*-Bu), 3.36 (1H, d, *J*=18 Hz, 2-H_α), 3.69 (1H, d, *J*=18 Hz, 2-H_β), 3.82 (3H, s, OCH₃), 4.50 (1H, d, *J*=5 Hz, 6-H), 5.00 (2H, s, CH₂O), 5.14 (1H, d, *J*=5 Hz, CH(NH)CO), 5.49 (1H, d, *J*=5 Hz, NHCOOC₄H₉), 6.01 (1H, dd, *J*=5 and 10 Hz, 7-H), 6.92 (1H, s, CHPh₂), 6.93 (2H, d, *J*=9 Hz, aromatic H), 6.99 (2H, d, *J*=9 Hz, aromatic H), 7.28~7.48 (14H, m, aromatic H), 7.63 (1H, d, *J*=10 Hz, CONH); FD-MS *m/z* 767 (M⁺).

Diphenylmethyl 7β-[D-α-(*tert*-Butoxycarbonylamino)-α-[4-(4-methoxybenzyl)oxyphenyl]acetamido]-3-ethoxycarbonylmethoxy-3-cephem-4-carboxylate 1β-Oxide (**4b**)

To a solution of **2** (1.0 g, 1.3 mm) in DMSO (8 ml) were added ethyl bromoacetate (**3b**) (435 mg, 2.61 mm) and *N,N*-diisopropylethylamine (252 mg, 1.96 mm) at room temp. After being stirred for 4 hours at the same temp, the reaction mixture was poured into 0.5% HCl (50 ml) under ice-cooling and extracted with EtOAc (100 ml). The extract was washed with brine (50 ml×2), dried (MgSO₄) and evaporated. The residue was purified by column chromatography on silica gel (eluent; benzene - acetone, 15:1~10:1), and crystallized from MeOH to give 420 mg (38.0%) of **4b**: MP 206~208°C;

Table 2. ^1H NMR and IR spectral data and yield of 4.

4

Compound			^1H NMR, δ (CDCl_3)					IR (KBr) cm^{-1}	Yield (%)
No.	X	R	2- H_α (1H, d, $J=17$ Hz)	2- H_β (1H, d, $J=17$ Hz)	6-H (1H, d, $J=5$ Hz)	7-H (1H, dd, $J=5, 9$ Hz)	Other protons		
4a	OBh	H	3.25	3.70	4.29	5.95	1.43 (9H, s), 3.80 (3H, s), 4.46 and 4.59 (2H, ABq, $J=16$ Hz), 4.98 (2H, s), 5.12 (1H, d, $J=6$ Hz), 5.56 (1H, d, $J=16$ Hz), 6.87 (1H, s), 6.91 (2H, d, $J=8$ Hz), 6.92 (1H, s), 6.97 (2H, d, $J=8$ Hz), 7.22~7.51 (25H, m)	1790, 1705, 1610	37.9
4c	O(<i>iso</i> -Pr)	H	3.41	3.85	4.47	6.00	1.22 (6H, d, $J=7$ Hz), 1.44 (9H, s), 3.82 (3H, s), 4.35 and 4.48 (2H, ABq, $J=16$ Hz), 5.00 (2H, s), 5.02 (1H, m), 5.13 (1H, d, $J=6$ Hz), 5.54 (1H, d, $J=6$ Hz), 6.93 (2H, d, $J=6$ Hz), 6.95 (1H, s), 6.98 (2H, d, $J=8$ Hz), 7.26~7.54 (15H, m)	1785, 1705, 1610	33.0
4d	O(<i>iso</i> -Bu)	H	3.40	3.83	4.46	6.00	0.90 (6H, d, $J=7$ Hz), 1.44 (9H, s), 1.90 (1H, m), 3.82 (3H, s), 3.88 (2H, d, $J=7$ Hz), 4.39 and 4.52 (2H, ABq, $J=16$ Hz), 5.00 (2H, s), 5.13 (1H, d, $J=6$ Hz), 5.55 (1H, d, $J=6$ Hz), 6.92 (2H, d, $J=8$ Hz), 6.95 (1H, s), 6.98 (2H, d, $J=8$ Hz), 7.26~7.54 (15H, m)	1785, 1710, 1610	33.5
4e	OCH_2Ph	H	3.33	3.77	4.37	5.98	1.44 (9H, s), 3.82 (3H, s), 4.42 and 4.55 (2H, ABq, $J=16$ Hz), 5.00 (2H, s), 5.12 (2H, s), 5.13 (1H, d, $J=6$ Hz), 5.56 (1H, d, $J=6$ Hz), 6.93 (2H, d, $J=8$ Hz), 6.94 (1H, s), 6.98 (2H, d, $J=8$ Hz), 7.24~7.52 (20H, m)	1790, 1700, 1610	36.5



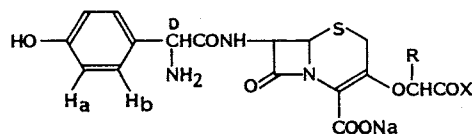
4f	OEt	CH ₃	3.28	3.95	4.44	5.98	1.20 (3H, t, <i>J</i> =7 Hz), 1.44 (9H, s), 1.47 (3H, s), 3.81 (3H, s), 4.04~4.20 (2H, m), 4.65 (1H, q, <i>J</i> =7 Hz), 4.99 (2H, s), 5.11 (1H, d, <i>J</i> =6 Hz), 5.47 (1H, d, <i>J</i> =6 Hz), 6.92 (2H, d, <i>J</i> =8 Hz), 6.96 (2H, d, <i>J</i> =8 Hz), 6.97 (1H, s), 7.24~7.52 (15H, m)	1785, 1695, 1620	13.8
4g	N(CH ₃) ₂	H	3.56	3.92	4.48	6.00	1.45 (9H, s), 2.66 (3H, s), 2.87 (3H, s), 3.82 (3H, s), 4.42 and 4.62 (2H, ABq, <i>J</i> =16 Hz), 5.00 (2H, s), 5.14 (1H, d, <i>J</i> =6 Hz), 5.55 (1H, d, <i>J</i> =6 Hz), 6.94 (2H, d, <i>J</i> =8 Hz), 6.97 (1H, s), 6.99 (2H, d, <i>J</i> =8 Hz), 7.26~7.52 (15H, m)	1785, 1715, 1660	30.0
4h	NEt ₂	H	3.59	3.94	4.47	5.99	1.01 (3H, t, <i>J</i> =7 Hz), 1.07 (3H, t, <i>J</i> =7 Hz), 1.43 (9H, s), 2.96 (2H, q, <i>J</i> =7 Hz), 3.29 (2H, q, <i>J</i> =7 Hz), 3.82 (3H, s), 4.40 and 4.59 (2H, ABq, <i>J</i> =14 Hz), 4.99 (2H, s), 5.11 (1H, d, <i>J</i> =6 Hz), 5.50 (1H, d, <i>J</i> =6 Hz), 6.92 (2H, d, <i>J</i> =8 Hz), 6.95 (1H, s), 6.98 (2H, d, <i>J</i> =8 Hz), 7.27~7.50 (14H, m), 7.62 (1H, d, <i>J</i> =9 Hz)	1790, 1725, 1675	40.4
4i		H	3.60	3.94	4.48	6.01	1.44 (9H, s), 3.04~3.68 (8H, m), 3.82 (3H, s), 4.40 and 4.59 (2H, ABq, <i>J</i> =16 Hz), 5.00 (2H, s), 5.13 (1H, d, <i>J</i> =6 Hz), 5.49 (1H, d, <i>J</i> =6 Hz), 6.93 (2H, d, <i>J</i> =8 Hz), 6.96 (1H, s), 6.98 (2H, d, <i>J</i> =8 Hz), 7.28~7.50 (15H, m)	1785, 1715, 1670	31.3
4j		H	3.40	3.74	4.58	5.88	1.44 (9H, s), 1.68~2.24 (4H, m), 2.98~3.22 (2H, m), 3.80 (3H, s), 4.36~4.63 (1H, m), 4.41 and 4.57 (2H, ABq, <i>J</i> =16 Hz), 5.00 (2H, s), 5.12 (1H, d, <i>J</i> =6 Hz), 5.53 (1H, d, <i>J</i> =6 Hz), 6.70 (1H, s), 6.92 (2H, d, <i>J</i> =8 Hz), 7.00 (1H, s), 7.00 (2H, d, <i>J</i> =8 Hz), 7.22~7.52 (25H, m)	1790, 1710, 1680	35.0

Table 3. ¹H NMR and IR spectral data and yield of 1.

1

Compound			¹ H NMR, δ (D ₂ O)							IR (KBr)	Yield
No.	X	R	H _a (2H, d, <i>J</i> =8 Hz)	H _b (2H, d, <i>J</i> =8 Hz)	6-H (1H, d, <i>J</i> =5 Hz)	7-H (1H, d, <i>J</i> =5 Hz)	2-H _{α} (1H, d, <i>J</i> =17 Hz)	2-H _{β} (1H, d, <i>J</i> =17 Hz)	Other protons	cm ⁻¹ β -Lactam	from 4 (%)
1a	ONa	H	6.95	7.37	5.09	5.51	3.21	3.46	4.30 and 4.40 (2H, ABq, <i>J</i> =16 Hz, OCH ₂ CO)	1750	62.2
1c	O(<i>iso</i> -Pr)	H	6.95	7.35	5.07	5.45	3.24	3.56	1.26 (6H, d, <i>J</i> =7 Hz, CH ₃ ×2), 4.53 and 4.63 (2H, ABq, <i>J</i> =17 Hz, OCH ₂ CO), 5.09 (1H, m, CH(CH ₃) ₂)	1750	58.6
1d	O(<i>iso</i> -Bu)	H	6.93	7.35	5.07	5.53	3.25	3.54	0.92 (6H, d, <i>J</i> =7 Hz, CH ₃ ×2), 1.97 (1H, m, CH(CH ₃) ₂), 4.01 (2H, d, <i>J</i> =7 Hz, CH ₂ CH(CH ₃) ₂), 4.60 and 4.71 (2H, ABq, OCH ₂ CO)	1750	58.0
1e	OCH ₂ Ph	H	6.92	7.33	4.93	5.51	3.14	3.42	4.58 and 4.68 (2H, ABq, <i>J</i> =17 Hz, OCH ₂ CO), 5.25 (2H, br s, CH ₂ Ph), 7.45 (5H, br s, Ph)	1745	57.3
1f	OEt	CH ₃	6.93	7.35	5.06	5.53	3.23	3.51	1.27 (3H, t, <i>J</i> =7 Hz, CH ₂ CH ₃), 1.49 (3H, d, <i>J</i> =7 Hz, CHCH ₃), 4.24 (2H, q, <i>J</i> =7 Hz, CH ₂ CH ₃), 4.71 (1H, q, <i>J</i> =7 Hz, CHCH ₃)	1745	61.5
1g	N(CH ₃) ₂	H	6.96	7.38	5.09	5.55	3.20	3.50	2.94 (3H, s, CH ₃), 2.98 (3H, s, CH ₃)	1760	59.4
1h	NEt ₂	H	6.94	7.34	5.07	5.51	3.21	3.47	1.10 (3H, t, <i>J</i> =7 Hz, CH ₂ CH ₃), 1.17 (3H, t, <i>J</i> =7 Hz, CH ₂ CH ₃), 3.31 (2H, q, <i>J</i> =7 Hz, CH ₂ CH ₃), 3.37 (2H, q, <i>J</i> =7 Hz, CH ₂ CH ₃), 4.71 and 4.78 (2H, ABq, <i>J</i> =16 Hz, OCH ₂ CO)	1755	56.0
1i		H	6.95	7.37	5.08	5.55	3.22	3.50	3.47~3.80 (8H, m, morpholine)	1755	61.8
1j		H	7.01	7.42	5.12, 5.13 (2×d)	5.56, 5.58 (2×d)	3.21, 3.23 (2×d)	3.55, 3.56 (2×d)	1.80~2.40 (4H, m, CH ₂ ×2), 3.45~3.62 (2H, m, NCH ₂), 4.23~4.38 (1H, m, NCHCH ₂)	1755	53.5

^a Racemization occurred in the process from 5j to 1j.

IR (KBr) cm^{-1} 1780, 1700, 1655; ^1H NMR (CDCl_3) δ 1.23 (3H, t, $J=7$ Hz, CH_2CH_3), 1.43 (9H, s, *tert*-Bu), 3.40 (1H, d, $J=17$ Hz, 2-H_α), 3.82 (3H, s, OCH_3), 3.83 (1H, d, $J=17$ Hz, 2-H_β), 4.16 (2H, q, $J=7$ Hz, CH_2CH_3), 4.36 and 4.51 (2H, ABq, $J=17$ Hz, OCH_2COO), 4.46 (1H, d, $J=5$ Hz, 6-H), 4.99 (2H, s, CH_2O), 5.13 (1H, d, $J=6$ Hz, $\text{CH}(\text{NH})\text{CO}$), 5.53 (1H, d, $J=6$ Hz, $\text{NHCOOC}_4\text{H}_9$), 6.00 (1H, dd, $J=5$ and 9 Hz, 7-H), 6.92 (2H, d, $J=8$ Hz, aromatic H), 6.95 (1H, s, CHPh_2), 6.98 (2H, d, $J=8$ Hz, aromatic H), 7.30~7.50 (15H, m, aromatic H and CONH); FD-MS m/z 853 (M^+);

Anal Calcd for $\text{C}_{45}\text{H}_{47}\text{N}_3\text{O}_{12}\text{S}$: C 63.29, H 5.55, N 4.92.

Found: C 63.35, H 5.56, N 5.10.

Similarly, compounds **4a** and **4c~4j** were prepared from **2** using the same procedure for **4b**. Their spectral data and yield are summarized in Table 2.

Diphenylmethyl 7 β -[D- α -(*tert*-Butoxycarbonylamino)- α -[4-(4-methoxybenzyl)oxyphenyl]acetamido]-3-ethoxycarbonylmethoxy-3-cephem-4-carboxylate (**5b**)

To a solution of **4b** (400 mg, 0.47 mm) in DMF (3.5 ml) was added dropwise phosphorus tribromide (127 mg, 0.47 mm) under ice-cooling. After being stirred for 30 minutes at the same temp, the reaction mixture was poured into H_2O (40 ml) and extracted with EtOAc (50 ml). The extract was washed with brine (40 ml), dried (MgSO_4) and evaporated under reduced pressure. The residue was purified by column chromatography on silica gel (eluent; benzene - acetone, 30:1~20:1) to give 303 mg (70.0%) of **5b** as an amorphous solid. IR (KBr) cm^{-1} 1780, 1705; ^1H NMR (CDCl_3) δ 1.24 (3H, t, $J=7$ Hz, OCH_2CH_3), 1.42 (9H, s, *tert*-Bu), 3.26 (1H, d, $J=16$ Hz, 2-H_α), 3.40 (1H, d, $J=16$ Hz, 2-H_β), 3.82 (3H, s, OCH_3), 4.18 (2H, q, $J=7$ Hz, OCH_2COO), 4.46 (2H, s, OCH_2COO), 4.98 (2H, s, CH_2O), 4.99 (1H, d, $J=5$ Hz, 6-H), 5.16 (1H, d, $J=6$ Hz, $\text{CH}(\text{NH})\text{CO}$), 5.59 (1H, d, $J=6$ Hz, $\text{NHCOOC}_4\text{H}_9$), 5.68 (1H, dd, $J=5$ and 9 Hz, 7-H), 6.56 (1H, d, $J=9$ Hz, CONH), 6.92 (2H, d, $J=9$ Hz, aromatic H), 6.93 (1H, s, CHPh_2), 6.97 (2H, d, $J=9$ Hz, aromatic H), 7.30~7.50 (14H, m, aromatic H); FD-MS m/z 838 ($\text{M}+1$) $^+$.

Sodium 7 β -[D- α -Amino- α -(4-hydroxyphenyl)acetamido]-3-ethoxycarbonylmethoxy-3-cephem-4-carboxylate (**1b**)

To a mixture of TFA (3.5 ml) and anisole (0.7 ml) was added **5b** (250 mg, 0.30 mm) under ice-cooling. After being stirred for 50 minutes at the same temp, the resulting solution was slowly added dropwise to a mixture of Et_2O and *n*-hexane (1:2, 30 ml). The precipitated trifluoroacetate of the desired product was collected by filtration, washed with a mixture of Et_2O and *n*-hexane (1:2, 30 ml). The above trifluoroacetate and NaHCO_3 (50 mg, 0.60 mm) were dissolved in H_2O (5 ml), and the solution was treated by column chromatography on Sephadex LH-20 (eluent; H_2O), and then lyophilized to give 120 mg (85.0%) of **1b** as white solid: IR (KBr) cm^{-1} 1750, 1670, 1600; ^1H NMR (D_2O) δ 1.27 (3H, t, $J=7$ Hz, OCH_2CH_3), 3.27 (1H, d, $J=17$ Hz, 2-H_α), 3.56 (1H, d, $J=17$ Hz, 2-H_β), 4.26 (2H, q, $J=7$ Hz, OCH_2CH_3), 4.56 and 4.67 (2H, ABq, $J=16$ Hz, OCH_2COO), 5.06 (1H, d, $J=5$ Hz, 6-H), 5.54 (1H, d, $J=5$ Hz, 7-H), 6.91 (2H, d, $J=8$ Hz, aromatic H), 7.34 (2H, d, $J=8$ Hz, aromatic H).

Compounds **1a** and **1c~1j** were similarly prepared from **4a** and **4c~4j** (Table 2) using the same procedures described for **1b**. Their spectral data and yield are summarized in Table 3.

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